Review

[Pulsed Field](https://doi.org/10.59958/hsf.7141) Ablation for the Treatment of Atrial Fibrillat[ion: A](https://journal.hsforum.com/) Review and a Look into its Future

Pavithran Guttipatti¹, Najla Saadallah¹, Elaine Y. Wan^{1,*}

¹Division of Cardiology, Department of Medicine, Vagelos College of Physicians and Surgeons, Columbia University, New York, NY 10032, USA *Correspondence: eyw2003@cumc.columbia.edu (Elaine Y. Wan)

Submitted: 18 December 2023 Revised: 11 January 2024 Accepted: 29 January 2024 Published: 22 February 2024

Abstract

Pulsed field ablation (PFA) is a novel technology to treat atrial fibrillation (AF) utilizing electric fields to induce nonthermal irreversible electroporation of electrically active cardiac tissue to induce cardiac cell death. PFA offers improved safety benefits compared to traditional radiofrequency ablation (RFA) and cryoablation by specifically ablating only cardiac tissue. However, there are avenues for further optimization including neurological risk associated with microbubble formation and left atrial function post ablation. Various PFA devices with different electric pulse waveforms have been studied and tested in human trials, with the majority utilizing microsecond duration pulses. Shorter nanosecond duration pulses, or nanosecond PFA, is beginning to be studied for AF ablation. In this review we will delve into current waveforms used for PFA, areas for improvement, mechanisms behind nanosecond PFA, and its clinical impact for cardiac ablation.

Keywords

arrhythmia; catheter ablation; atrial fibrillation; pulsed field ablation; nanosecond; electroporation

Introduction

AF is the most common arrhythmia seen in clinical practice, which left uncontrolled can lead to devastating consequences such as stroke and heart failure [1]. In patients refractory to rate control or anti-arrhythmic medication, catheter ablation of the electrically active tissue around the pulmonary veins can reduce arrhythmia burden [2]. The main modes of ablation in clinical practice to[da](#page-7-0)y are radiofrequency ablation (RFA) or cryoablation, which rely on thermal heating or cooling to destroy tissue. PFA is an emerging technology that is a non-thermal ablation [m](#page-7-1)ethod that relies on creating permanent pores in the cell membrane of cardiomyocytes [3–5]. PFA may be safer than RFA with lower risk of complications such as esophageal lesions, pulmonary vein stenosis, and nerve injury, in part due to the specificity of PFA for ablating cardiomyocytes over other cell types [6–8]. The parameters of the electric pulse utilized have an important role in the effect of PFA on tissue. This review will discuss the waveforms utilized by current PFA devices tested clinically, and dive into the emerging research o[n u](#page-8-0)[ltr](#page-8-1)a-short nanosecond duration PFA.

PFA Waveform Parameters

An electric field has multiple properties that can be varied to cause differential effects on tissue. Electric stimulation is applied in a pulsed manner usually in the form of a square wave, but other waveforms such as a sinusoidal pattern are also possible [3]. The stimulation can be a monophasic configuration where voltage pulses are continuously positive, or a biphasic configuration where the voltage pulse has a positive region and a negative region [9,10]. The amplitude of the pulse can [be](#page-7-2) altered to change the field strength, quantified in Volts (V)/meter (m), and the duration of the pulse can be altered by changing the pulse width [11]. A pulse train can be formed by modifying the inter-pu[ls](#page-8-2)[e in](#page-8-3)terval or frequency of pulses to determine the total number of pulses delivered to the tissue $[10]$. Further characteristics such as the positioning of the electrode from which the [fiel](#page-8-4)d is emitted and distance to the tissue can also alter the effects on the target [12]. Current PFA devices in clinical testing and use typically employ micr[osec](#page-8-3)ond-scale pulse widths (Fig. 1A); nanosecond PFA entails utilizing much shorter pulse widths at the expense of greater voltages to perform ablation (Fig. [1B](#page-8-5)). The details of current clinical devices will be discussed first, followed by review of research into the [mo](#page-1-0)re nascent nanosecond domain.

Current PFA Clinical Devices and their Waveforms

A PFA delivery system requires a generator that can produce the pulse waveform as well as a catheter with electrodes for application of electric fields. There are multiple PFA devices currently on the market, each utilizing their own specified pulse parameters and, in most cases, their own custom catheters [7,8,13–16] (Table 1, Ref. [7,8,13– 19]). Although the exact details of the pulse waveform used by these devices is proprietary, all of these devices

Publisher's Note: Forum Multimedia Publishing stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Fig. 1. Schematic depictions of PFA waveforms (A) A biphasic waveform with two pulses of microsecond duration (B) A biphasic waveform with shorter, nanosecond duration pulses. The voltage is increased due to the shorter pulse width. PFA, Pulsed field ablation.

are similar in that the pulse width of stimulation is in the microsecond or greater order of magnitude. Initial studies tested both monophasic and biphasic waveforms [7]. However, monophasic waveforms have been shown to trigger greater skeletal muscle contraction requiring general anesthesia, while biphasic waveforms carry the advantage of reduced contraction and ability to perform ablat[io](#page-8-6)n under conscious sedation [7,20].

The initial trial of PFA in 2018 on 22 patients with paroxysmal AF (pAF) tested both endocardial and epicardial ablation using [a](#page-8-6) [pul](#page-8-7)sed waveform of millisecond duration, delivered in trains over a few seconds [17]. The voltage was the only varied parameter, with endocardial ablation using 900–1000 V and epicardial ablation using 2100–2400 V. In a subsequent larger study of 81 patients in the IMPULSE and PEFCAT trials, t[he a](#page-8-8)uthors performed pulmonary vein isolation (PVI) using shorter microsecond-scale pulses delivered in a bipolar fashion with both monophasic and biphasic waveforms tested [7]. Voltages in this study were between 900 to 1000 V for monophasic and 1800 to 2000 V for biphasic waveforms respectively. The authors further optimized the biphasic waveform, although the specifics were not able to be [dis](#page-8-6)closed due to proprietary information. PFA was delivered using the Farapulse™ System, from Farapulse Inc. (Menlo Park, CA, USA) now acquired by Boston Scientific Inc., which utilizes a pentaspline catheter deployable in a flower or basket configuration. Extending this approach to persistent AF (persAF), the PersAFOne trial carried out PFA for PVI as well as left atrial posterior wall ablation [18]. The waveforms in this trial also employed microsecond-scale biphasic pulses at 1600 to 2000 V.

Based on these first in human studies, Schmidt *et al*. [19] described the adoption and usage of PFA te[chn](#page-8-9)ology in Germany in an all-comer group of 191 AF patients in the 5S study. Utilizing the Farapulse™ ablation system approved in Europe, they followed the biphasic microsecond[scal](#page-8-10)e protocol of previous trials. Voltage was also kept at 1800 to 2000 V and a train of 5 consecutive waveforms was delivered for a total application time of 2.5 seconds. A survey of 24 centers in Europe that adopted PFA after Farapulse™'s approval covered 1758 patients with a mixture of pAF and persAF, and also utilized the same standard waveform protocol [21].

Other PFA devices have also been developed with a different catheter shape and their own individualized waveform. The Multi-Channel PFA Generator from Biosense Webster Inc. [\(Irv](#page-8-11)ine, CA, USA) is compatible with the electroanatomical mapping system CARTO3. In the in-SPIRE trial this system was used to treat 226 patients with pAF, employing a square wave biphasic waveform with microsecond-width pulses at 1800 V delivered in trains for a total 250 milliseconds per application [13]. A sphere-shaped lattice-tipped catheter allowing for toggling between PFA, RFA, and electroanatomic mapping, the Sphere-9 developed by Affera Inc. (Watertown, MA, USA) which was then acquired by Medtronic (Minneapolis, [MN](#page-8-12), USA), has also been studied for ablation of pAF or persAF [14]. This device also delivers biphasic microsecondwidth pulses, in a train of 3–5 seconds. Notably the catheter also irrigates the delivery area with saline. In a study of 178 patients with pAF or persAF, various waveforms were devel[ope](#page-8-13)d to attempt to increase electrical isolation efficiency [14]. The first evolution PULSE1 used 3–5.5 second trains of lesions with saline irrigation at 4–30 mL/min, the second-generation PULSE2 used 4 second trains with saline at 15 mL/min, and the final generation PULSE3 also used 4 [sec](#page-8-13)ond trains and 15 mL/min. The exact parameters of the waveform and the difference between PULSE2 and PULSE3 is kept proprietary. However, these waveform characteristics have a great impact on tissue effects, as the durability of PVI at an average of 3 months post-ablation improved from 32% of patients with all pulmonary veins still isolated with PULSE1 to 64% with PULSE2 and to 90% with PULSE3.

Further evidence for the importance of waveform parameters comes from the ECLIPSE-AF study of the CEN-TAURI system for PFA ablation, developed by Galvanize Therapeutics Inc. (San Carlos, CA, USA) [15]. The CEN-TAURI system has a generator that is compatible with three

| Catheter structure | Study | Pulse width |
|------------------------------------|---------------------------------|-------------------|
| Pentaspline catheter deployable | Reddy <i>et al.</i> 2018 [17] | Initial studies: |
| in flower or basket-shaped con- | Reddy et al. 2019 [7] | Millisecond scale |
| figuration | Reddy <i>et al.</i> 2020 [18] | Refined waveform: |
| | Schmidt et al. 2022 [19] | Microsecond scale |
| Variable loop circular catheter | Duytschaever et al. 2023 [13] | Microsecond scale |
| | | |
| Spherical lattice-tipped catheter | Reddy <i>et al.</i> 2023 [14] | Microsecond scale |
| | | |
| Generator that is compatible with | Anić et al. 2023 [15] | Millisecond scale |
| existing commercial ablation | | |
| catheters | | |
| Annular electrode array catheter | Verma <i>et al.</i> 2023 [8] | Undisclosed pulse |
| | | width |
| Spherical electrode array catheter | Turagam <i>et al.</i> 2023 [16] | Microsecond scale |
| | | |

Table 1. Current PFA devices undergoing clinical testing or use.

different commercially available ablation catheters (Abbott TactiCath SE, Boston Scientific StablePoint, and Biosense Webster ThermoCool ST). As the workflow was optimized through the trial, patients ended up in 5 cohorts with different waveform settings or catheters used in each group. Waveforms ranged from 1.4 to 3.4 milliseconds duration, currents from 19 Amps to 25 Amps, and lesion diameter was varied. Similar to the Sphere-9 trial, the first two cohorts had low 90-day PVI durability while later cohorts with optimized parameters showed increased durability. Medtronic has also developed a catheter with a circular array of electrodes, the PulseSelect PFA System, tested in the PULSED AF trial [8]. Ablation is delivered in biphasic pulse trains of 100–200 milliseconds, with a voltage around 1500 V. The exact pulse width in the train is not disclosed. Finally, the Globe is a spherical electrode array PFA catheter from Kardium [I](#page-8-1)nc. (Burnaby, British Columbia, Canada) that was tested for PVI [16]. Interestingly, in this study patients received microsecond-width pulses at 1700 V in either a single application or three applications repetitively at the same site. Repetitive application led to more durable PVI, with 30% of singlea[ppl](#page-8-15)ication patients showing persistent complete PVI at 2–3 months versus 100% in the multiple application group. Therefore, in addition to the individual parameters of each pulse, the total amount of applications can also be varied to achieve the desired outcome. While a single application would be most time efficient, an ablation method that has greater safety but requires multiple applications may be worthwhile. The clinical studies discussed above using PFA with microsecond or greater duration pulses have shown promising safety data and may lower the risk of complications seen with conventional ablation such as pulmonary vein stenosis and esophageal lesions [8]. However, there are still areas where PFA's safety profile may be improved.

Microbubble For[ma](#page-8-15)tion and Neurological Risk during PFA

Ablation procedures have a low rate of neurological complications such as stroke, however studies have shown a significant portion of patients show silent neurological findings on magnetic resonance imaging (MRI) after ablation that may be attributable to air or thrombus embolization [22]. MRI findings can be categorized as silent cerebral events (SCE) that are diffusion-weighted imaging (DWI) positive, or silent cerebral lesions (SCL) that are DWI positive as well as fluid-attenuated inverse recovery sequence ([FL](#page-8-16)AIR) positive [23]. While SCLs have more evidence associated with histological changes and scar formation, the FLAIR changes cannot be detected until 2–7 days post procedure [24]. SCEs are a more sensitive marker that only rely on DWI, andt[hus](#page-8-17) are around three times as common as SCLs, however their pathological significance is less clear [23]. The rate of SCE with traditional RFA in the literature is aroun[d 6](#page-8-18).8–24% and for SCL between 7.4–8.3%, and appears to vary with the ablation technology used [23]. For example, cryoballoon ablation shows lower rates of SCE [betw](#page-8-17)een 8.9–18% and SCL between 4.3–5.6% [23]. This raises the question of how PFA fares compared to current technology, and whether the waveform parameter[s ut](#page-8-17)ilized may alter the incidence of neurological lesions.

In the IMPULSE and PEFCAT trials the a[utho](#page-8-17)rs typically observed ultrasonic microbubbles on intracardiac echocardiography immediately after PFA application, hypothesized to be due to electrolysis of water in the presence of an electric field [7]. However, they demonstrate that no patients presented with evidence of stroke, transient ischemic attack (TIA), or systemic embolism, and additionally performed MRIs on 13 patients that did not show evidence of silent ischemic e[ve](#page-8-6)nts. A further combined analy-

sis with the results of the PEFCATII trial showed 16 out of 18 patients showed no MRI lesions after PFA; one patient with a history of TIA's experienced brief dysphasia after the procedure with a small correlate lesion found on DWI but not FLAIR SCE, and a second patient also showed a DWI positive lesion SCE [25]. The 5S study also carried out MRI in a subset of 53 patients, 24–48 hours after the ablation. In the initial validation stage of this trial, the PFA catheter's electrograms were verified with a circular mapping catheter, requiring cat[hete](#page-8-19)r exchange. After the first 25 patients the study transitioned to the streamline phase where the circular mapping catheter was omitted. During the validation stage two patients developed strokes after the PFA procedure: the first with blurred vision and novel ischemic lesion on MRI, and the second with right hand weakness and gait ataxia with multiple small acute lesions in the basal ganglia and cerebellum. In both patients the symptoms had resolved at later follow-up. During the streamline phase no patients had neurological symptoms, suggesting that the catheter exchange may have played a role. However, diffusion weighted MRI showed asymptomatic cerebral injury in 10/53 (19%) of patients albeit without neurological deficits [19].

An alternate PFA system developed by Biosense Webster Inc. in the inSPIRE trial also underwent testing for brain ischemia using MRI [13]. In this study an initial 4 [out](#page-8-10) of 6 patients showed SCLs, leading to workflow modifications such as instating a 10 second pause between PFA applications, minimizing catheter exchanges, and ensuring anticoagulation adherence. [Fur](#page-8-12)ther results showed SCL in 4 out of 33 patients (12%). The authors note all the lesions were asymptomatic and resolved by a 3 month follow up. The Sphere-9 system that combines PFA, RFA, and electroanatomic mapping showed SCEs in 5 (9.8%) and SCLs in 3 (5.9%) of 51 patients in their initial study [18]. A second larger study of the device showed SCEs in 7 (7.9%) and SCLs in 6 (6.7%) out of 89 patients [14]. With Medtronic's circular catheter, the PulseSelect™ system tested in the PULSED AF trial, 1 out of 150 pAF pat[ien](#page-8-9)ts and 0 out of 150 persAF patients developed stroke post PFA [8]. Further, MRI studies of a 45 patient [co](#page-8-13)hort showed 4 (8.9%) had postprocedural SCLs while SCEs were not reported. Functional testing of this cohort was also carried out with Mini Mental State Examinations, which did not signifi[can](#page-8-1)tly differ from before PFA to 30 day follow up.

Interestingly in the ECLIPSE-AF trial testing the CENTAURI PFA system, monitoring for microbubbles with intracardiac echocardiography during the index procedures did not demonstrate any microbubbles in 61 procedures [15]. Furthermore, MRI of 36 patients showed SCE in 4 (11.4%) and SCL in 0 (0%) of patients. The four patients with SCEs were all in cohort 4 of the study where the Boston Scientific Inc. StablePoint catheter was used. Simila[r to](#page-8-14) other studies all patients were asymptomatic, and lesions resolved at 30 day follow up. Finally, the PULSE-EU study of the Kardium Inc. Globe system that tested single versus multiple applications of the same PFA waveform showed SCEs in 2 patients (28.6%) in the single application group and 1 patient (11.1%) in the multiple application group [16]. This suggests additional PFA applications do not increase the number of brain lesions, although sample sizes are small.

The overall rate of clinically symptomatic neurological de[fici](#page-8-15)ts after PFA appears to be low, as confirmed by the EU-PORIA real-world outcomes study of the Farapulse system in Europe, which finds TIA or stroke in 7 out of 1233 patients (0.6%) [26]. The only randomized controlled trial of PFA recently compared the Farapulse system from Boston Scientific against traditional RFA or cryoballoon ablation for pAF and demonstrated non-inferiority in efficacy as well as seriou[s ad](#page-8-20)verse events at 1 year [27]. In this study, there were no clinical strokes and 1 TIA seen in the 305 patients treated with PFA, while there was 1 stroke and no TIAs seen in the 302 patients treated with thermal ablation. Thus, there appears to be a low rate and in[com](#page-8-21)parable difference in symptomatic neurological deficits from these different ablation modalities. A brain MRI on a subset of patients however showed 3/33 (9%) treated with PFA had asymptomatic ischemic phenomena (SCE or SCL) compared to 0/37 treated with thermal ablation. It is unclear whether these asymptomatic neurological lesions are associated with long-term effects on cognition [23]. Regardless a method of ablation that could minimize the formation of these lesions would be ideal. The lack of any microbubbles or SCLs with the CENTAURI system compared to other PFA systems suggests that there are param[eter](#page-8-17)s that can be modified to prevent their development.

Left Atrial Function Post-Ablation with PFA

The left atrium (LA) has important functions that are perturbed in AF and that may be differentially altered by ablation modality. The LA's hemodynamic function can be divided into reservoir, conduit, and booster phases [28]. The LA acts as a reservoir during ventricular systole, enlarging without increasing chamber pressure to prevent pulmonary hypertension. It acts as a conduit between the pulmonary veins and the left ventricle (LV) during vent[ricu](#page-8-22)lar diastole, and its contraction at end-diastole boosts LV filling. Furthermore, the LA also plays a neurohormonal role, with atrial stretch triggering local natriuretic peptide release, as well as nerve activation leading to inhibition of central sympathetic outflow [29]. AF is associated with deleterious structural and functional changes including LA enlargement and reduced atrial systolic function [30,31]. Prolonged AF leads to progressive LA fibrosis that reduces LA capacitance and thus reser[voi](#page-8-23)r function [32].

The effect of conventional ablation on LA function has conflicting results in the literature. Mechanistically restoration of sinus rhythm via ablation may boost systolic function, while the creation of scar tissue from ablation may counteract this benefit and lead to reduced atrial distensibility. Electron beam tomography on pAF patients after RFA revealed left atrial edema in a large portion of patients [33]. Furthermore, the severity of the edema depended on the extent and amount of radiofrequency energy delivered, although the edema naturally resolved with time. Long term follow-up of AF patients after RFA via echocardiography [has](#page-8-25) shown improvement in LA size as well as LV diastolic and systolic function [34]. On the other hand, contrast enhanced computed tomography (CECT) study of pAF patients by Lemola *et al*. [35] demonstrated that LA systolic function, measured via LA emptying fraction (LAEF), declines after ablation. [In](#page-8-26) contrast, Verma *et al*. [36] performed echocardiography and cine electron beam computed tomography (EBCT) on [pA](#page-8-27)F and persAF patients after pulmonary vein antrum isolation and noted no adverse effect on LA function and even improvement in LAEF at [6 m](#page-9-0)onths post-ablation. The authors note to have a larger sample of patients than Lemola and colleagues [36]. However, a later study by Wylie and colleagues [37] that used cardiovascular magnetic resonance imaging (CMR) of pAF and persAF patients revealed lower LAEF after ablation. Furthermore, the authors studied patients w[ho](#page-9-0) underwent limited pulmonary vein ostial ablation, [and](#page-9-1) surmised that more extensive circumferential ablation with linear lesions to the LA posterior wall, roof and/or mitral isthmus may have an even greater effect on function. Given the numerous conflicting studies, a meta-analysis of the effect of catheter ablation found that while left atrial size is improved, LAEF decreases in pAF patients and is not significantly altered in persAF patients [38]. Therefore, conventional ablation appears to have a detrimental effect on LA systolic function in at least a subset of patients [29]. RFA may also in rare cases cause LA diastolic dysfunction severe enough to lead to pulmonary hy[pert](#page-9-2)ension and dyspnea, known as stiff LA syndrome [29,39]. This complication occurred in 1.4% of patients in one study and was [mo](#page-8-23)re likely to occur in patients with severe LA scarring [39].

The c[lini](#page-8-23)[cal](#page-9-3) significance of reduced atrial function is not well established. However, reduced LA contractile function is predictive of AF r[ecu](#page-9-3)rrence after catheter ablation [40,41]. While it is unclear if improving LA function after ablation will lead to greater long-term success of cardioversion, methods of ablation that preserve LA function would be ideal. Initial results comparing PFA with RFA and cryoab[lati](#page-9-4)[on](#page-9-5) showed less late gadolinium enhancement at 3 months post-ablation, suggesting less chronic fibrosis, and greater recovery of LA reservoir and booster pump function with PFA [42]. Further work will be needed to confirm these results in a wider patient pool, as well as study the effect of different pulse parameters on LA function. Nanosecond PFA that allows for reduced total energy delivery to the tissue may better preserve LA function and reduce the risk of complications such as stiff LA syndrome. More studies need to be performed to evaluate this.

Is Nanosecond PFA The Future?

Pulsed electric fields have been extensively studied in non-cardiac cells for applications ranging from laboratory transfection to tumor cell killing [43]. The mechanistic findings from the wider body of non-cardiac research is presented first, followed by insights from cardiac specific studies.

The principal mechanism behind [PF](#page-9-7)A is electroporation of the cell membrane, leading to the formation of pores that allow for free flow in and out of the cell and apoptosis [44,45]. In order for pore formation to occur, the magnitude of the electric field across the cell membrane must reach a certain strength [46]. The magnitude of the electric field also determines whether the pores are formed transien[tly,](#page-9-8) [le](#page-9-9)ading to reversible electroporation that is commonly used in laboratories for transfection, or permanently, leading to irreversible e[lect](#page-9-10)roporation and cell death [46]. Microsecond and millisecond duration electric field pulses lead to movement of ions in the extracellular and intracellular space, with the cell membrane acting as a barrier that leads to capacitive charging. This phenom[eno](#page-9-10)n amplifies the externally applied electric field, known as the Maxwell-Wagner effect, such that the critical magnitude needed across the cell membrane for pore formation is reached [47,48]. Pores have been observed to preferentially form at the poles of cells that face electrodes since they experience the greatest ion alignment [48–50].

Shorter nanosecond-duration pulsed electric fields (nsPEF) ar[e a](#page-9-11)[lso](#page-9-12) capable of inducing cell death [51–53]. Nanosecond fields are too brief to allow for capacitive charging to amplify the external electricf[ield](#page-9-12) [ac](#page-9-13)ross the cell membrane [52,54]. Modeling and experimental results demonstrate that nanosecond electric pulses [ar](#page-9-14)e [ca](#page-9-15)pable of charging the smaller intracellular membranes of organelles, allowing for electroporation of structures such as the nucleus, [mito](#page-9-16)[ch](#page-9-17)ondria, and endoplasmic reticulum, which may contribute to apoptosis [55]. Furthermore, it was shown that electroporation of intracellular membranes could occur without electroporation of the outer cell membrane in COS-7 cells using nsPEF [56]. However, followup studies on a wider range of cell typ[es s](#page-9-18)howed that nsPEF are indeed capable of causing pore formation on the plasma membrane [57–60]. The pores formed from nsPEF may be smaller in size than the pores from [m](#page-9-19)icrosecond and millisecond duration fields, at around 1–1.5 nanometers, and thus not detectable in experiments by large dyes such as propidium [iod](#page-9-20)i[de](#page-9-21) that could not pass through them [61]. Nanosecond electric fields thus appear capable of creating a

transmembrane potential that is enough to electroporate the cell membrane, but due to the lack of amplification from capacitive charging require a higher external field strength to do so [48,62]. Although a higher voltage is used, the shorter duration of the pulse means the total energy delivered is often lower with nsPEF. Pores also form more uniformly across the cell rather than at the poles [48,54].

Puls[ed](#page-9-12) [elec](#page-9-23)tric fields may also lead to cell death via migration of membrane phosphatidylserine, a phagocytic signal on apoptotic cells, from the cytosolic to the extracellular side of the membrane via pores. [Thi](#page-9-12)[s p](#page-9-17)rocess has been demonstrated to occur in response to both microsecond and nanosecond duration fields [63]. Comparison of 10 nanosecond, 300 nanosecond, 1.8 microsecond, and 9 microsecond pulses in cells showed that the 10 nanosecond pulses were the least efficient at triggering cell death, but more selective between [tw](#page-9-24)o different cell types [64]. While the 1.8 microsecond pulses led to pores large enough for propidium iodide uptake, the 10 nanosecond pulses did not, however both pulses resulted in phosphatidylserine externalization. Nanosecond fields may thus have t[he a](#page-9-25)dvantage of even greater selectivity than microsecond fields, although more pulse applications may be required for effective ablation given the lower efficiency of cell killing. In addition to nanopore formation of the cell membrane and organelles and phosphatidylserine externalization, other mechanisms of action for nsPEF have also been described. Membrane cholesterol concentration and lipid rafts appear to be an important determinant of nsPEF response, as is oxidation of membrane lipids [65,66]. Further research is needed on these effects of nsPEF and comparison with microsecond and millisecond fields.

The above-described insights on nsPEF action come from the wide body of literature studying th[eir](#page-9-26) [eff](#page-9-27)ect on non-cardiac cell types. For application to AF ablation, it is important to understand the mechanistic effects of nsPEF on cardiac cells. Nanosecond pulsed fields have been shown to similarly affect cardiomyocytes by creating pores in the sarcolemma, allowing for calcium flow independent of ion channels [67]. Paralleling non-cardiac cell results, nsPEF requires greater voltages to achieve electroporation in cardiomyocytes compared to microsecond fields. In a detailed study on isolated mouse, pig, and rabbit cardiomyocytes, Semenov [and](#page-9-28) colleagues compared 10 microsecond, 800 nanosecond, and 200 nanosecond width shocks [68]. The authors initially identified the voltage that was necessary to trigger calcium transients in the cell for each pulse duration, then used five times that voltage for electroporation. At the 5 *×* voltage the nanosecond pulses de[mon](#page-10-0)strated significantly reduced electroporation compared to the microsecond pulses. The authors note this benefit of nsPEF for defibrillation, which would reduce unnecessary electroporation and cardiac damage. However, for the purposes of ablation, this suggests that increased voltage parameters may be necessary to achieve the same degree of electroporation with nsPEF. It is worth noting that electroporation was determined in this study by the uptake of propidium iodide, a membrane-impermeable dye that fluoresces red upon binding nuclear material. However, this dye molecule may be too large to enter and detect pores that are exceedingly small such as nanopores formed by nsPEF. Exposure of rat embryonic cardiomyocytes to 10 nanosecond versus 4 millisecond duration fields showed that at pulse parameters that led to the same level of calcium influx, the millisecond fields led to more propidium iodide uptake, suggesting that millisecond fields create larger membrane pores while nsPEF creates smaller pores [48]. Furthermore, calcium imaging revealed influx beginning at the poles of the cells with millisecond fields, while nsPEF led to homogenous influx across the cell. This supports results discussed earlier in other cells that nsPEF leads [to](#page-9-12) more uniform electroporation across an individual cell.

A key feature of cardiomyocytes is their rod-shaped structure. Pulse width has interestingly been shown to determine whether the orientation of cardiomyocytes within the electric field affects electroporation. In nanosecond duration pulses, cardiomyocytes that are oriented perpendicular to the field are more electroporated than cells oriented parallel [69]. However, in the microsecond pulse domain cell orientation does not have an effect on electroporation efficacy. Furthermore, at even longer millisecond pulse durations cells oriented parallel to the field are more electroporatedt[han](#page-10-1) those perpendicular. These results suggest that while nsPEF cause pore formation uniformly across a cell, on a tissue level cells that are oriented perpendicular to the applied field will be more likely to be targeted. Microsecond pulses may carry the advantage of more thorough ablation in tissue with differently oriented cells. However, with advancements in understanding of the myofiber architecture in the atrium the orientation selectivity of nsPEF may become useful for ablating specific tracts [70]. Additionally, catheters may be designed to apply multidirectional electric fields if complete ablation of all orientations is desired. It is worth noting these studies were carried out in H9c2 rat ventricular myoblasts and AC16 [hum](#page-10-2)an ventricular myocytes in culture $[69]$. Given the complex threedimensional nature of the atrium and factors such as extracellular matrix, vessels, and nerves that alter structure, further studies will be needed to confirm this phenomenon *in vivo*, not just in cell cultur[e.](#page-10-1)

Non-Cardiac Clinical Studies of Nanosecond PFA

While nanosecond pulsed fields have not yet been studied in humans for cardiac ablation, they are beginning to be studied in the field of oncology. Different duration pulsed fields including millisecond, microsecond, and nanosecond fields have all been researched for tumor ablation and have been reviewed previously [71–73]. Mi-

crosecond pulses have been applied for direct killing of tumor cells via irreversible electroporation, as well as in electrochemotherapy where the electric field is used to increase tumor cell permeability to chemotherapeutic drugs. Nanosecond pulses are an emerging area within tumor treatment as well. While microsecond duration pulses in tumors appear to lead to cell death via necrosis, nanosecond duration pulses may favor apoptosis, possibly due to smaller pore size formation that only allows ion fluxes while maintaining adenosine tri-phosphate (ATP) that is necessary for apoptosis [74].

The first human trial of nsPEF was carried out for basal cell carcinoma and ablated 10 lesions in three patients [75]. Electric field pulses 100 nanoseconds wide were delivered at a[n a](#page-10-3)mplitude of 30,000 V/cm. The results demonstrated that 7 of the 10 treated lesions were completely devoid of basaloid cells, two partially regressed, and only one l[esi](#page-10-4)on recurred as squamous cell carcinoma by week ten. The treatment led to edema and crust formation, followed by the gradual return of normal skin appearance. Importantly, no visible scars were observed at the treated sites. Initial trials in humans have also demonstrated the feasibility of nsPEF for the treatment of other cutaneous lesions such as seborrheic keratosis and sebaceous gland hyperplasia [76,77]. Clinical studies on applications for visceral organs are limited; however, Liu and colleagues [78] have trialed nsPEF for treating hepatocellular carcinoma (HCC) in 15 patients [79]. Using 300 nanosecond duration pulses at 30,[000](#page-10-5) [V t](#page-10-6)he authors were able to ablate liver tumors within 0.5 cm of critical structures such as the portal [vein](#page-10-7), hepatic vein, biliary tree, or gastrointestinal tract. These results are encouraging [fo](#page-10-8)r translation to cardiac applications.

Cardiac Studies of Nanosecond PFA

Although there have been limited human trials of nsPEF for treatment of tumors, there are no clinical studies applying nsPEF to the heart. Current insights are limited to research performed in pre-clinical animal models. Xie and colleagues [80] applied nsPEF to rabbit hearts in an *ex-vivo* Langendorff perfusion setup. The nsPEF were delivered via two electrodes that were inserted transmurally into the ventricle, and the waveform used a pulse width of 350 nanoseconds ap[plie](#page-10-9)d in different pulse trains and amplitudes. Optical mapping was carried out to assess the effects on heart conduction, and successfully confirmed that electrical block could be achieved consistently at 2300 V when electrodes were 2.3 mm apart and 4000 V when 4 millimeters apart. A single pulse was not sufficient for ablation and multiple pulses in a train were needed. Notably the authors state that their nanosecond ablation approach uses a thousand times less energy than the microsecond pulses in the seminal PFA studies of Lavee and colleagues [44]. The nanosecond pulses use higher field strengths but at a much lower duration. Further studies from the same group have shown efficacy of nsPEF at 10,000-12,000 V for ablation in pig atria and ventricular myocardium with transmural penetration, albeit with a surgical approach for electrode positioning [81]. Catheter-based delivery would be the primary modality of ablation in clinical practice. There has been initial work on nanosecond PFA delivery utilizing a circular multielectrode catheter, the CellFX system from Pulse Biosciences [Inc](#page-10-10). (Hayward, CA, USA), for ablation in pigs [82]. Ablation was attempted at the superior vena cava, right superior pulmonary vein, and discrete sites on the atria, with successful delivery at 22/22 sites and histology showing transmural cell necrosis in 21 of 22 lesions. These d[ata](#page-10-11) were published in a recent abstract and more detailed findings will be insightful.

Nanosecond PFA has also begun to be studied for ventricular targets. Tan *et al*. [83] utilized the CellFX generator to create nanosecond electric fields, but delivered the stimulation using conventional transvenous pacemaker leads, either with a single lead in a bipolar manner from helix-to-ring, or using two l[ead](#page-10-12)s. In their study they targeted ablation to the right ventricular septum of canines, and observed that a single lead was limited in the amount of electric field energy that could be delivered due to arcing, resulting in only mild reduction in cardiac electrogram voltages and no evidence of lesion creation on cardiac MRI or histopathology. The two-lead system allowed for a stronger electric field delivery and showed significant electrogram amplitude decreases, along with associated late gadolinium enhancement on MRI, dense fibrosis, and fewer than 5% residual cardiomyocytes on histology. Interestingly the authors also compared the lesions from nanosecond PFA with RFA on histopathology, and found the RFA lesions showed more necrotic cardiomyocytes interspersed with fibrosis and hemorrhage. This data fits with nanosecond fields promoting apoptosis over necrosis as discussed earlier, and may benefit LA function after ablation. However, the study in pig atria from Koruth *et al*. [82] described transmural cell necrosis at nanosecond PFA sites, which suggests that other parameters besides pulse duration may play a role. Tan and colleagues [83] also note that one of their nanosecond PFA cases showed adhere[nt o](#page-10-11)rganizing thrombus formation at the ablation site. This may be due to the screw-in nature of the lead delivering PEF rather than the nanosecond fields, but is a potent[ial r](#page-10-12)isk. Nanosecond fields have also been shown to be capable of transiently blocking the atrioventricular (AV) conduction system in canine hearts when ablation is targeted to the interventricular septum [84]. This effect is dependent on the level of energy delivered and is reversible with time, with histology demonstrating ablation of myocardium and sparing of Purkinje fibers. It is unclear if the transient blocking of the AV conduct[ion](#page-10-13) system is specific to nanosecond versus microsecond pulses, but is a point of caution when applying pulsed fields to the ventricle during ablation.

Future Directions

Pulsed field ablation has burst onto the electrophysiology stage as an effective method of ablation for AF that is safer than conventional RFA or cryoablation. While there are multiple PFA devices in clinical testing, each with their individual proprietary waveforms, all of them utilize microsecond or greater duration pulses. While generating nanosecond duration electric field pulses was challenging in the past due to their ultrashort and high voltage nature, technological advancements have allowed for nsPEF to become studied more easily [85]. Nanosecond fields differ mechanistically from the microsecond and millisecond fields that have currently been tested in human patients. By creating smaller nanopores in the cell membrane as well as intracellular organelles nsP[EF](#page-10-14) may favor apoptosis over necrosis, and also allow for myofiber orientation specific ablation. Initial results from preclinical models suggest that nanosecond PFA can be a promising treatment for AF. Nanosecond PFA may allow lower energy application that could improve the safety profile of ablation. While the results discussed show that nanosecond PFA can ablate cardiac tissue in porcine, canine, and rabbit models there have not been long-term studies on the durability of electrical isolation. These studies would need to be conducted to confirm the irreversibility of nanosecond PFA. The selectivity of nanosecond pulses for myocardium and sparing of nerves and esophageal tissue also needs to be demonstrated. If confirmed this would open the avenue for studies directly comparing nanosecond versus microsecond PFA for AF ablation in animal models or humans. Other potential barriers to implementation in practice include the number of applications of electric field pulses necessary to successfully ablate tissue with nanosecond versus microsecond fields, which could affect procedure time and time under anesthesia. It remains to be seen if nsPEF leads to lower rates of microbubble formation and cerebral ischemic findings. The impact of nsPEF versus longer pulse durations on LA function after ablation would also benefit from study. Other outcomes such as atrial arrhythmia recurrence could also be compared with different pulse duration parameters. In sum, nanosecond PFA is an area for further exploration with potential for improving the surgical treatment of atrial fibrillation.

Availability of Data and Materials

No original data were required for the preparation of this review.

Author Contributions

EYW and PG contributed to the design of this work. PG and NS drafted the initial manuscript. PG and EYW edited and all authors read and approved the final manuscript. All authors agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All authors contributed to editorial changes in the manuscript.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

Not applicable.

Funding

P.G. is supported by the Dean's Research Fellowship of Columbia University. EYW is supported by NIH R01 HL152236.

Conflict of Interest

E.Y.W. has been a consultant for Boston Scientific and national PI of Boston Scientific sponsored LUX-DX Trends trial. The authors have no financial interest in the devices described in the manuscript.

References

- [1] Lippi G, Sanchis-Gomar F, Cervellin G. Global epidemiology of atrial fibrillation: An increasing epidemic and public health challenge. International Journal of Stroke: Official Journal of the International Stroke Society. 2021; 16: 217–221.
- [2] Haïssaguerre M, Jaïs P, Shah DC, Takahashi A, Hocini M, Quiniou G, *et al*. Spontaneous initiation of atrial fibrillation by ectopic beats originating in the pulmonary veins. The New England Journal of Medicine. 1998; 339: 659–666.
- [3] Verma A, Asivatham SJ, Deneke T, Castellvi Q, Neal RE, 2nd. Primer on Pulsed Electrical Field Ablation: Understanding the Benefits and Limitations. Circulation. Arrhythmia and Electrophysiology. 2021; 14: e010086.
- [4] Schaack D, Schmidt B, Tohoku S, Bordignon S, Urbanek L, Ebrahimi R, *et al*. Pulsed Field Ablation for Atrial Fibrillation. Arrhythmia & Electrophysiology Review. 2023; 12: e11.
- [5] Di Monaco A, Vitulano N, Troisi F, Quadrini F, Romanazzi I, Calvi V, *et al*. Pulsed Field Ablation to Treat Atrial Fibrillation: A Review of the Literature. Journal of Cardiovascular Development and Disease. 2022; 9: 94.
- [6] Stewart MT, Haines DE, Miklavčič D, Kos B, Kirchhof N, Barka N, *et al*. Safety and chronic lesion characterization of pulsed field ablation in a Porcine model. Journal of Cardiovascular Electrophysiology. 2021; 32: 958–969.
- [7] Reddy VY, Neuzil P, Koruth JS, Petru J, Funosako M, Cochet H, *et al*. Pulsed Field Ablation for Pulmonary Vein Isolation in Atrial Fibrillation. Journal of the American College of Cardiology. 2019; 74: 315–326.
- [8] Verma A, Haines DE, Boersma LV, Sood N, Natale A, Marchlinski FE, *et al*. Pulsed Field Ablation for the Treatment of Atrial Fibrillation: PULSED AF Pivotal Trial. Circulation. 2023; 147: 1422–1432.
- [9] Ye X, Liu S, Yin H, He Q, Xue Z, Lu C, *et al*. Study on Optimal Parameter and Target for Pulsed-Field Ablation of Atrial Fibrillation. Frontiers in Cardiovascular Medicine. 2021; 8: 690092.
- [10] García-Sánchez T, Amorós-Figueras G, Jorge E, Campos MC, Maor E, Guerra JM, *et al*. Parametric Study of Pulsed Field Ablation With Biphasic Waveforms in an *In Vivo* Heart Model: The Role of Frequency. Circulation. Arrhythmia and Electrophysiology. 2022; 15: e010992.
- [11] Meckes D, Emami M, Fong I, Lau DH, Sanders P. Pulsed-field ablation: Computational modeling of electric fields for lesion depth analysis. Heart Rhythm O2. 2022; 3: 433–440.
- [12] Howard B, Verma A, Tzou WS, Mattison L, Kos B, Miklavčič D, *et al*. Effects of Electrode-Tissue Proximity on Cardiac Lesion Formation Using Pulsed Field Ablation. Circulation. Arrhythmia and Electrophysiology. 2022; 15: e011110.
- [13] Duytschaever M, De Potter T, Grimaldi M, Anic A, Vijgen J, Neuzil P, *et al*. Paroxysmal Atrial Fibrillation Ablation Using a Novel Variable-Loop Biphasic Pulsed Field Ablation Catheter Integrated With a 3-Dimensional Mapping System: 1-Year Outcomes of the Multicenter inspIRE Study. Circulation. Arrhythmia and Electrophysiology. 2023; 16: e011780.
- [14] Reddy VY, Peichl P, Anter E, Rackauskas G, Petru J, Funasako M, *et al*. A Focal Ablation Catheter Toggling Between Radiofrequency and Pulsed Field Energy to Treat Atrial Fibrillation. JACC. Clinical Electrophysiology. 2023; 9: 1786–1801.
- [15] Anić A, Phlips T, Brešković T, Koopman P, Girouard S, Mediratta V, *et al*. Pulsed field ablation using focal contact forcesensing catheters for treatment of atrial fibrillation: acute and 90-day invasive remapping results. Europace: European Pacing, Arrhythmias, and Cardiac Electrophysiology: Journal of the Working Groups on Cardiac Pacing, Arrhythmias, and Cardiac Cellular Electrophysiology of the European Society of Cardiology. 2023; 25: euad147.
- [16] Turagam MK, Neuzil P, Petru J, Funasako M, Koruth JS, Reinders D, *et al*. PV Isolation Using a Spherical Array PFA Catheter: Application Repetition and Lesion Durability (PULSE-EU Study). JACC. Clinical Electrophysiology. 2023; 9: 638–648.
- [17] Reddy VY, Koruth J, Jais P, Petru J, Timko F, Skalsky I, *et al*. Ablation of Atrial Fibrillation With Pulsed Electric Fields: An Ultra-Rapid, Tissue-Selective Modality for Cardiac Ablation. JACC. Clinical Electrophysiology. 2018; 4: 987–995.
- [18] Reddy VY, Anic A, Koruth J, Petru J, Funasako M, Minami K, *et al*. Pulsed Field Ablation in Patients With Persistent Atrial Fibrillation. Journal of the American College of Cardiology. 2020; 76: 1068–1080.
- [19] Schmidt B, Bordignon S, Tohoku S, Chen S, Bologna F, Urbanek L, *et al*. 5S Study: Safe and Simple Single Shot Pulmonary Vein Isolation With Pulsed Field Ablation Using Sedation. Circulation. Arrhythmia and Electrophysiology. 2022; 15: e010817.
- [20] Koruth J, Kuroki K, Iwasawa J, Enomoto Y, Viswanathan R, Brose R, *et al*. Preclinical Evaluation of Pulsed Field Ablation: Electrophysiological and Histological Assessment of Thoracic Vein Isolation. Circulation. Arrhythmia and Electrophysiology.

2019; 12: e007781.

- [21] Ekanem E, Reddy VY, Schmidt B, Reichlin T, Neven K, Metzner A, *et al*. Multi-national survey on the methods, efficacy, and safety on the post-approval clinical use of pulsed field ablation (MANIFEST-PF). Europace: European Pacing, Arrhythmias, and Cardiac Electrophysiology: Journal of the Working Groups on Cardiac Pacing, Arrhythmias, and Cardiac Cellular Electrophysiology of the European Society of Cardiology. 2022; 24: 1256–1266.
- [22] Deneke T, Shin DI, Balta O, Bünz K, Fassbender F, Mügge A, *et al*. Postablation asymptomatic cerebral lesions: long-term follow-up using magnetic resonance imaging. Heart Rhythm. 2011; 8: 1705–1711.
- [23] Deneke T, Jais P, Scaglione M, Schmitt R, DI Biase L, Christopoulos G, *et al*. Silent cerebral events/lesions related to atrial fibrillation ablation: a clinical review. Journal of Cardiovascular Electrophysiology. 2015; 26: 455–463.
- [24] Deneke T, Nentwich K, Krug J, Müller P, Grewe PH, Mügge A, *et al*. Silent Cerebral Events after Atrial Fibrillation Ablation - Overview and Current Data. Journal of Atrial Fibrillation. 2014; 6: 996.
- [25] Reddy VY, Dukkipati SR, Neuzil P, Anic A, Petru J, Funasako M, *et al*. Pulsed Field Ablation of Paroxysmal Atrial Fibrillation: 1-Year Outcomes of IMPULSE, PEFCAT, and PEFCAT II. JACC. Clinical Electrophysiology. 2021; 7: 614–627.
- [26] Schmidt B, Bordignon S, Neven K, Reichlin T, Blaauw Y, Hansen J, *et al*. EUropean real-world outcomes with Pulsed field ablatiOn in patients with symptomatic atRIAl fibrillation: lessons from the multi-centre EU-PORIA registry. Europace: European Pacing, Arrhythmias, and Cardiac Electrophysiology: Journal of the Working Groups on Cardiac Pacing, Arrhythmias, and Cardiac Cellular Electrophysiology of the European Society of Cardiology. 2023; 25: euad185.
- [27] Reddy VY, Gerstenfeld EP, Natale A, Whang W, Cuoco FA, Patel C, *et al*. Pulsed Field or Conventional Thermal Ablation for Paroxysmal Atrial Fibrillation. The New England Journal of Medicine. 2023; 389: 1660–1671.
- [28] Mehrzad R, Rajab M, Spodick DH. The three integrated phases of left atrial macrophysiology and their interactions. International Journal of Molecular Sciences. 2014; 15: 15146–15160.
- [29] Packer M. Effect of catheter ablation on pre-existing abnormalities of left atrial systolic, diastolic, and neurohormonal functions in patients with chronic heart failure and atrial fibrillation. European Heart Journal. 2019; 40: 1873–1879.
- [30] Gupta DK, Shah AM, Giugliano RP, Ruff CT, Antman EM, Grip LT, *et al*. Left atrial structure and function in atrial fibrillation: ENGAGE AF-TIMI 48. European Heart Journal. 2014; 35: 1457–1465.
- [31] Allessie M, Ausma J, Schotten U. Electrical, contractile and structural remodeling during atrial fibrillation. Cardiovascular Research. 2002; 54: 230–246.
- [32] Habibi M, Lima JAC, Khurram IM, Zimmerman SL, Zipunnikov V, Fukumoto K, *et al*. Association of left atrial function and left atrial enhancement in patients with atrial fibrillation: cardiac magnetic resonance study. Circulation. Cardiovascular Imaging. 2015; 8: e002769.
- [33] Okada T, Yamada T, Murakami Y, Yoshida N, Ninomiya Y, Shimizu T, *et al*. Prevalence and severity of left atrial edema detected by electron beam tomography early after pulmonary vein ablation. Journal of the American College of Cardiology. 2007; 49: 1436–1442.
- [34] Reant P, Lafitte S, Jaïs P, Serri K, Weerasooriya R, Hocini M, *et al*. Reverse remodeling of the left cardiac chambers after catheter ablation after 1 year in a series of patients with isolated atrial fibrillation. Circulation. 2005; 112: 2896–2903.
- [35] Lemola K, Desjardins B, Sneider M, Case I, Chugh A, Good

E, *et al*. Effect of left atrial circumferential ablation for atrial fibrillation on left atrial transport function. Heart Rhythm. 2005; 2: 923–928.

- [36] Verma A, Kilicaslan F, Adams JR, Hao S, Beheiry S, Minor S, *et al*. Extensive ablation during pulmonary vein antrum isolation has no adverse impact on left atrial function: an echocardiography and cine computed tomography analysis. Journal of Cardiovascular Electrophysiology. 2006; 17: 741–746.
- [37] Wylie JV, Jr, Peters DC, Essebag V, Manning WJ, Josephson ME, Hauser TH. Left atrial function and scar after catheter ablation of atrial fibrillation. Heart Rhythm. 2008; 5: 656–662.
- [38] Xiong B, Li D, Wang J, Gyawali L, Jing J, Su L. The Effect of Catheter Ablation on Left Atrial Size and Function for Patients with Atrial Fibrillation: An Updated Meta-Analysis. PLoS ONE. 2015; 10: e0129274.
- [39] Gibson DN, Di Biase L, Mohanty P, Patel JD, Bai R, Sanchez J, *et al*. Stiff left atrial syndrome after catheter ablation for atrial fibrillation: clinical characterization, prevalence, and predictors. Heart Rhythm. 2011; 8: 1364–1371.
- [40] Wen S, Indrabhinduwat M, Brady PA, Pislaru C, Miller FA, Ammash NM, *et al*. Post Procedural Peak Left Atrial Contraction Strain Predicts Recurrence of Arrhythmia after Catheter Ablation of Atrial Fibrillation. Cardiovascular Ultrasound. 2021; 19: 22.
- [41] Khan HR, Yakupoglu HY, Kralj-Hans I, Haldar S, Bahrami T, Clague J, *et al*. Left Atrial Function Predicts Atrial Arrhythmia Recurrence Following Ablation of Long-Standing Persistent Atrial Fibrillation. Circulation. Cardiovascular Imaging. 2023; 16: e015352.
- [42] Nakatani Y, Sridi-Cheniti S, Cheniti G, Ramirez FD, Goujeau C, André C, *et al*. Pulsed field ablation prevents chronic atrial fibrotic changes and restrictive mechanics after catheter ablation for atrial fibrillation. Europace: European Pacing, Arrhythmias, and Cardiac Electrophysiology: Journal of the Working Groups on Cardiac Pacing, Arrhythmias, and Cardiac Cellular Electrophysiology of the European Society of Cardiology. 2021; 23: 1767–1776.
- [43] Rubinsky B, Onik G, Mikus P. Irreversible electroporation: a new ablation modality–clinical implications. Technology in Cancer Research & Treatment. 2007; 6: 37–48.
- [44] Lavee J, Onik G, Mikus P, Rubinsky B. A novel nonthermal energy source for surgical epicardial atrial ablation: irreversible electroporation. The Heart Surgery Forum. 2007; 10: E162– E167.
- [45] Davalos RV, Mir ILM, Rubinsky B. Tissue ablation with irreversible electroporation. Annals of Biomedical Engineering. 2005; 33: 223–231.
- [46] Kotnik T, Rems L, Tarek M, Miklavčič D. Membrane Electroporation and Electropermeabilization: Mechanisms and Models. Annual Review of Biophysics. 2019; 48: 63–91.
- [47] Kotnik T, Miklavcic D. Second-order model of membrane electric field induced by alternating external electric fields. IEEE Transactions on Bio-medical Engineering. 2000; 47: 1074– 1081.
- [48] Semenov I, Zemlin C, Pakhomova ON, Xiao S, Pakhomov AG. Diffuse, non-polar electropermeabilization and reduced propidium uptake distinguish the effect of nanosecond electric pulses. Biochimica et Biophysica Acta. 2015; 1848: 2118–2125.
- [49] Tekle E, Astumian RD, Chock PB. Selective and asymmetric molecular transport across electroporated cell membranes. Proceedings of the National Academy of Sciences of the United States of America. 1994; 91: 11512–11516.
- [50] Kotnik T, Pucihar G, Miklavcic D. Induced transmembrane voltage and its correlation with electroporation-mediated molecular transport. The Journal of Membrane Biology. 2010; 236: 3–13.
- [51] Beebe SJ, Fox PM, Rec LJ, Somers K, Stark RH, Schoenbach

KH. Nanosecond pulsed electric field (nsPEF) effects on cells and tissues: apoptosis induction and tumor growth inhibition. IEEE Transactions on Plasma Science. 2002; 30: 286–292.

- [52] Beebe SJ, Fox PM, Rec LJ, Willis ELK, Schoenbach KH. Nanosecond, high-intensity pulsed electric fields induce apoptosis in human cells. FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology. 2003; 17: 1493–1495.
- [53] Stacey M, Stickley J, Fox P, Statler V, Schoenbach K, Beebe SJ, *et al*. Differential effects in cells exposed to ultra-short, high intensity electric fields: cell survival, DNA damage, and cell cycle analysis. Mutation Research. 2003; 542: 65–75.
- [54] Gowrishankar TR, Weaver JC. Electrical behavior and pore accumulation in a multicellular model for conventional and supraelectroporation. Biochemical and Biophysical Research Communications. 2006; 349: 643–653.
- [55] Schoenbachk KH, Joshi RP, Kolb JF, Chen N, Stacey M, Buescher ES, *et al*. Ultrashort electrical pulses open a new gateway into biological cells (pp. 205–209). In Conference Record of the Twenty-Sixth International Power Modulator Symposium. 2004 and 2004 High-Voltage Workshop. 2004.
- [56] Tekle E, Oubrahim H, Dzekunov SM, Kolb JF, Schoenbach KH, Chock PB. Selective field effects on intracellular vacuoles and vesicle membranes with nanosecond electric pulses. Biophysical Journal. 2005; 89: 274–284.
- [57] Ibey BL, Roth CC, Pakhomov AG, Bernhard JA, Wilmink GJ, Pakhomova ON. Dose-dependent thresholds of 10-ns electric pulse induced plasma membrane disruption and cytotoxicity in multiple cell lines. PLoS ONE. 2011; 6: e15642.
- [58] Ibey BL, Xiao S, Schoenbach KH, Murphy MR, Pakhomov AG. Plasma membrane permeabilization by 60- and 600-ns electric pulses is determined by the absorbed dose. Bioelectromagnetics. 2009; 30: 92–99.
- [59] Pakhomov AG, Kolb JF, White JA, Joshi RP, Xiao S, Schoenbach KH. Long-lasting plasma membrane permeabilization in mammalian cells by nanosecond pulsed electric field (nsPEF). Bioelectromagnetics. 2007; 28: 655–663.
- [60] Deng J, Schoenbach KH, Buescher ES, Hair PS, Fox PM, Beebe SJ. The effects of intense submicrosecond electrical pulses on cells. Biophysical Journal. 2003; 84: 2709–2714.
- [61] Pakhomov AG, Bowman AM, Ibey BL, Andre FM, Pakhomova ON, Schoenbach KH. Lipid nanopores can form a stable, ion channel-like conduction pathway in cell membrane. Biochemical and Biophysical Research Communications. 2009; 385: 181–186.
- [62] Son RS, Smith KC, Gowrishankar TR, Vernier PT, Weaver JC. Basic features of a cell electroporation model: illustrative behavior for two very different pulses. The Journal of Membrane Biology. 2014; 247: 1209–1228.
- [63] Vernier PT, Sun Y, Gundersen MA. Nanoelectropulse-driven membrane perturbation and small molecule permeabilization. BMC Cell Biology. 2006; 7: 37.
- [64] Ibey BL, Pakhomov AG, Gregory BW, Khorokhorina VA, Roth CC, Rassokhin MA, *et al*. Selective cytotoxicity of intense nanosecond-duration electric pulses in mammalian cells. Biochimica et Biophysica Acta. 2010; 1800: 1210–1219.
- [65] Cantu JC, Tarango M, Beier HT, Ibey BL. The biological response of cells to nanosecond pulsed electric fields is dependent on plasma membrane cholesterol. Biochimica et Biophysica Acta. 2016; 1858: 2636–2646.
- [66] Vernier PT, Levine ZA, Wu YH, Joubert V, Ziegler MJ, Mir LM, *et al*. Electroporating fields target oxidatively damaged areas in the cell membrane. PloS One. 2009; 4: e7966.
- [67] Wang S, Chen J, Chen MT, Vernier PT, Gundersen MA, Valderrábano M. Cardiac myocyte excitation by ultrashort high-field pulses. Biophysical Journal. 2009; 96: 1640–1648.
- [68] Semenov I, Grigoryev S, Neuber JU, Zemlin CW, Pakhomova ON, Casciola M, *et al*. Excitation and injury of adult ventricular cardiomyocytes by nano- to millisecond electric shocks. Scientific Reports. 2018; 8: 8233.
- [69] Dermol-Černe J, Batista Napotnik T, Reberšek M, Miklavčič D. Short microsecond pulses achieve homogeneous electroporation of elongated biological cells irrespective of their orientation in electric field. Scientific Reports. 2020; 10: 9149.
- [70] Pashakhanloo F, Herzka DA, Ashikaga H, Mori S, Gai N, Bluemke DA, *et al*. Myofiber Architecture of the Human Atria as Revealed by Submillimeter Diffusion Tensor Imaging. Circulation. Arrhythmia and Electrophysiology. 2016; 9: e004133.
- [71] Nuccitelli R. Application of Pulsed Electric Fields to Cancer Therapy. Bioelectricity. 2019; 1: 30–34.
- [72] Breton M, Mir LM. Microsecond and nanosecond electric pulses in cancer treatments. Bioelectromagnetics. 2012; 33: 106–123.
- [73] Campana LG, Daud A, Lancellotti F, Arroyo JP, Davalos RV, Di Prata C, *et al*. Pulsed Electric Fields in Oncology: A Snapshot of Current Clinical Practices and Research Directions from the 4th World Congress of Electroporation. Cancers. 2023; 15: 3340.
- [74] Esser AT, Smith KC, Gowrishankar TR, Weaver JC. Towards solid tumor treatment by nanosecond pulsed electric fields. Technology in Cancer Research & Treatment. 2009; 8: 289–306.
- [75] Nuccitelli R, Wood R, Kreis M, Athos B, Huynh J, Lui K, *et al*. First-in-human trial of nanoelectroablation therapy for basal cell carcinoma: proof of method. Experimental Dermatology. 2014; 23: 135–137.
- [76] Hruza GJ, Zelickson BD, Selim MM, Rohrer TE, Newman J, Park H, *et al*. Safety and Efficacy of Nanosecond Pulsed Electric Field Treatment of Seborrheic Keratoses. Dermatologic Surgery: Official Publication for American Society for Dermatologic Surgery [et Al.]. 2020; 46: 1183–1189.
- [77] Munavalli GS, Zelickson BD, Selim MM, Kilmer SL, Rohrer TE, Newman J, *et al*. Safety and Efficacy of Nanosecond Pulsed Electric Field Treatment of Sebaceous Gland Hyperplasia. Der-

matologic Surgery: Official Publication for American Society for Dermatologic Surgery [et Al.]. 2020; 46: 803–809.

- [78] Liu J, Fang C, Jin X, Tian G, Sun Z, Hong L, *et al*. Nanosecond pulsed electric field ablation-induced modulation of sphingolipid metabolism is associated with $Ly6c2⁺$ mononuclear phagocyte differentiation in liver cancer. Molecular Oncology. 2023; 17: 1093–1111.
- [79] Xu M, Xu D, Dong G, Ren Z, Zhang W, Aji T, *et al*. The Safety and Efficacy of Nanosecond Pulsed Electric Field in Patients With Hepatocellular Carcinoma: A Prospective Phase 1 Clinical Study Protocol. Frontiers in Oncology. 2022; 12: 869316.
- [80] Xie F, Varghese F, Pakhomov AG, Semenov I, Xiao S, Philpott J, *et al*. Ablation of Myocardial Tissue With Nanosecond Pulsed Electric Fields. PloS One. 2015; 10: e0144833.
- [81] Varghese F, Philpott JM, Neuber JU, Hargrave B, Zemlin CW. Surgical Ablation of Cardiac Tissue with Nanosecond Pulsed Electric Fields in Swine. Cardiovascular Engineering and Technology. 2023; 14: 52–59.
- [82] Koruth JS, Kawamura I, Maan A, Musikantow DR, Turagam MK, Miller MA, *et al*. PO-04-067 nanosecond pulsed field ablation: demonstration of halo-shaped lesions with a novel multielectrode system: initial preclinical experience. Heart Rhythm. 2023; 20: S516–S517.
- [83] Tan NY, Ladas TP, Christopoulos G, Sugrue AM, van Zyl M, Ladejobi AO, *et al*. Ventricular nanosecond pulsed electric field delivery using active fixation leads: a proof-of-concept preclinical study. Journal of Interventional Cardiac Electrophysiology: an International Journal of Arrhythmias and Pacing. 2022. (online ahead of print)
- [84] van Zyl M, Ladejobi AO, Tri JA, Yasin OZ, Connolly RJ, Danitz DJ, *et al*. Reversible Atrioventricular Conduction Impairment Following Bipolar Nanosecond Electroporation of the Interventricular Septum. JACC. Clinical Electrophysiology. 2021; 7: 255–257.
- [85] Sundararajan R. Nanosecond electroporation: another look. Molecular Biotechnology. 2009; 41: 69–82.